Table II. Localized MNDO Geometry

atom paira	dist, Å	bond order	two-center energy, eV	
1, 12	1.4743	1.0477	-16.698	
4, 12	1.3722	1.8111	-22.867	
1, 11	1.5231	0.9631	-14.717	
4, 5	1.5143	0.9697	-15.200	
1, 2	1.5345	0.9430	-13.012	
3, 4	1.5447	0.9515	-14.114	
2, 3	1.5402	0.9737	-14.641	
10, 11	1.5653	0.9744	-13.875	
5, 6	1.5806	0.9587	-13.841	
1, 9	1.6969	0.8128	-8.143	
4, 7	2.1814	0.0458	+0.254	

<sup>&</sup>lt;sup>a</sup> For atom numbering, see Table I.

cule, 11 SCF theory is not capable of giving an accurate description of the delocalized form and at least a simple  $2 \times 2$  configuration interaction (CI) involving the highest occupied (HOMO) and lowest unoccupied (LUMO) molecular orbitals is necessary.

## Results

Initial MNDO-SCF calculations on compounds 3 indicated that only the localized structures are minima on the potential energy surface. The delocalized structure 3b was only optimized by imposing the necessary symmetry constraints. This structure is a transition structure between the two localized forms and is 3.22 kcal/mol higher in energy. The MNDO-based  $2 \times 2$  CI results also give the localized forms as minima but now the symmetric form is also a minimum. The calculated  $\Delta H_{\rm f}$  was 108.245 kcal/mol for the localized form and 98.877 kcal/mol for the symmetric form. More importantly, the symmetric form is 10.37 kcal/mol lower in energy than the localized forms. The geometries of the CI-optimized structures are given in Tables I and II.

Like the MNDO calculations the AM1-CI results indicated the symmetric form as the minimum-energy form. However, neither the AM1-SCF method nor the AM1 2 × 2–CI method was able to find a stable localized structure. The geometry of the AM1 results is given in Table I and the calculated  $\Delta H_f$  is 117.049 kcal/mol. As a test of the AM1 procedure and for comparison with experimental results, calculations were done on semibullvalene itself. These results indicate the localized forms ( $\Delta H_f(AM1)$ ) = 83.303 kcal/mol) are the minimum-energy forms with the symmetric geometry ( $\Delta H_{\rm f}({\rm AM1}) = 87.470 \; {\rm kcal/mol}$ ) being a transition structure. The AM1 barrier of 4.17 kcal agrees well with the previous MNDO calculation of 5.7 kcal and the experimental results of 4.8 kcal.<sup>11</sup>

The question that remains is whether or not the predicted symmetric double-annelated semibullvalene is homoaromatic. It is true that the ring strain and anti-Bredt character of the localized geometries will cause them to be of high energy. However, there must be some interaction causing the molecule to adopt a symmetric geometry and this may be homoaromatic stabilization. The degree of stabilization (or destabilization) associated with a particular interaction can be indicated by using the energy partitioning from the MOPAC program into one-center (atomic) and two-center (bond) terms. 18 As can be seen (Tables I and II) typical stabilizations associated with the semibullvalene type single bonds (e.g., 2-3) are approximtely -14 eV. Although the 1-9 and 4-7 interactions are much weaker it is apparent that in the symmetric (homoaromatic) system 3b they are a stabilizing effect. For the localized form found in the MNDO calculations the

1-9 interaction is very stabilizing (corresponding to a single bond) and the 4-7 interaction is now destabilizing. It should be pointed out that the computed bond orders also indicate a favorable 1-9 and 4-7 interaction in the symmetric molecule. We, therefore, conclude that not only does the bisannelated semibullvalene 3 display appreciable homoconjugation but also that this homoconjugation is energy lowering and, therefore, 3 joins the elite ranks as a rare example of a neutral homoaromatic (ground state) hydrocarbon.

The disagreement between the MNDO and AM1 methods on the existence of a stable localized structure for the double-annelated semibullvalene 3a is of some concern. To answer this question properly would require a more complete search of the potential energy surface in order to locate transition structures and other possible minima. Currently, this is not feasible with our version of the MOPAC program. We are, however, interested in exploring the differences between AM1 and MNDO and continuing our studies in this area by encorporating new searching routines in the MOPAC program.

Further studies are also under way using ab initio methods. It has been shown previously that to get qualitative information on other homoaromatic systems via ab inito methods, it may be necessary to use very good basis sets (including polarization function) and also to include electron correlation. 19,20 The inclusion of correlation at the second-order Moller-Plessett (MP2) level has been shown to have a dramatic effect on systems of this type.

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## A Stereospecific Synthesis of 3-Aminodeoxystatine<sup>1</sup>

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Pepstatin (1), first isolated by Umezawa and co-workers in 1970,<sup>2</sup> is a pentapeptide that exhibits extremely potent inhibitory activity with a majority of the enzymatic family of aspartyl proteinases (for example, a  $K_i$  of 4.6 × 10<sup>-11</sup> M vs pepsin<sup>3</sup>). Mechanistically, 1 derives its activity from

the unusual amino acid statine (2), (3S,4S)-4-amino-3hydroxy-6-methylheptanoic acid, which functions either as a "transition state" 4 or "collected substrate" 5 insert

<sup>(19)</sup> Haddon, R. C.; Raghavachari, K. J. Am. Chem. Soc. 1983, 105, 118

<sup>(20)</sup> Haddon, R. C. J. Am. Chem. Soc. 1988, 110, 1108.

<sup>(1)</sup> Presented, in part, at the 20th National Medicinal Chemistry Symposium, Chapel Hill, NC, June 18, 1986, Abstract #34.
(2) Umezawa, H.; Aoyagi, T.; Morishima, H.; Matsuzaki, M.; Takeuchi, T. J. Antibiot. 1970, 23, 259.

<sup>(3)</sup> Marciniszyn, J.; Hartsuck, J. A.; Tang, J. J. Biol. Chem. 1976, 251,

<sup>(4)</sup> Workman, R. J.; Burkitt, D. W. Arch. Biochem. Biophys. 1979, 194,

Figure 1. Statine (2) and isosteric analogues 3 and 4.

Scheme I (only a series stereochemistry shown)

<sup>a</sup> Reaction conditions: (i) NH<sub>2</sub>OCH<sub>3</sub>·HCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>; (ii) triphenylphosphine, DEAD, THF; (iii) MsCl, Pyr, then K<sub>2</sub>CO<sub>3</sub>, acetone; (iv) Na, NH<sub>3</sub>, THF; (v) KOH, MeOH, H<sub>2</sub>O; (vi) 1 M K<sub>2</sub>CO<sub>3</sub>, CBZC1.

based on its structural analogy to the metastable intermediate proposed in the amide bond hydrolysis mechanism of aspartyl proteinases. Of particular interest within this class has been the enzyme renin, a member of the reninangiotensin cascade, whose inhibition may offer a useful treatment for human hypertension.6

This concept of transition-state analogy<sup>7</sup> has prompted a great deal of research in an effort to determine which electronic and structural parameters are important in designing effective statine analogues.8 Our intent was the design and stereospecific synthesis of isosterically modified statine residues that may shed light on the importance of hydrogen-bonding and electrostatic interactions in this series of enzymatic inhibitors.9 This note reports the stereospecific synthesis of two such analogues, (3S,4S)- and (3R,4S)-3-aminodeoxystatine (3 and 4), respectively<sup>10</sup> (Figure 1).

While literature procedures exist for the synthesis of β-amino acids, 11 we desired a stereochemically defined synthesis from starting materials of known absolute configuration which would eliminate the need for diastereo-

(5) Rich, D. H. J. Med. Chem. 1985, 28, 263

(7) Pauling, L. Chem. Eng. News 1946, 24, 1375. Leinhard, G. E.

Science Washington, D.C. 1973, 180, 149.

(9) At physiological pH (7.4), the  $\beta$  amino group would be expected to be protonated, thus allowing a potential measurement of the effect of an electrostatic interaction with the negatively charged aspartyl residues; see: James, M. N. G.; Sielecki, A. R. Biochemistry 1985, 24, 3701.

Chem. 1982, 231, 185 and references cited therein.

Scheme IIa

<sup>a</sup>Reaction conditions: (i) DBU, CH<sub>3</sub>I, CH<sub>3</sub>CN; (ii) TBSOTf, CH<sub>2</sub>Cl<sub>2</sub>, then saturated NaHCO<sub>3</sub>.

meric separations. This approach relied on the inversion of the  $\beta$ -hydroxyl of **5a** and **5b**, readily available through literature procedures, 12,13 with a nitrogen nucleophile that could easily be converted to a suitably protected form, as in 9a. We chose to utilize the stereospecific intramolecular Mitsunobu<sup>14</sup> approach developed by Miller et al.<sup>15</sup> to provide the desired stereochemistry at the  $\beta$  position as shown in Scheme I. In this variant of the Mitsunobu reaction a  $\beta$ -hydroxy-O-alkyl hydroxamate is cyclized wherein the O-alkyl hydroxamate moiety functions as the internal nucleophile and the resulting product is a functionalized  $\beta$ -lactam as shown in Scheme I. This reaction succeeds because the  $pK_a$  of the O-alkyl hydroxamate group (p $K_a = 9-10$ ) is below that of the conjugate base of dicarbethoxyhydrazide (threshold p $K_a \leq 13$ ).<sup>15</sup>

Condensation of 5a with O-methylhydroxylamine hydrochloride gave the crystalline O-methyl hydroxamate  $\mathbf{6a}$ in 58% yield. Intramolecular Mitsunobu cyclization of 6a was conducted by utilizing the standard reagents<sup>16</sup> to deliver the N-methoxy- $\beta$ -lactam 7a in 70% yield after chromatography on silica gel. Reductive removal of the amide methoxy<sup>17</sup> gave 8a, which was subjected to aqueous potassium hydroxide to cleave the  $\beta$ -lactam ring and neutralized and the free amino moiety was protected as the benzyloxycarbonyl (CBZ) urethane. The crude acid 9a was most conveniently purified by the formation of the crystalline dicyclohexylamine (DCHA) salt 10a. This series of reactions thus resulted in the synthesis of 3 in a suitably protected form that allowed further synthetic manipulation.

With the 3S diaster eomer 9a in hand, the sequence was repeated to prepare the 3R isomer 9b. Boc-statine (5b) was straightforwardly converted to the O-methyl hydroxamate 6b; however, the cyclization conditions of DEAD/triphenylphosphine were replaced with the twostep procedure developed by Floyd et al.<sup>17</sup> The chromatographic removal of the byproducts of the Mitsunobu reaction, triphenylphosphine and dicarboethoxyhydrazide. had become exceedingly difficult on a large scale; thus the procedure of  $\beta$ -mesylation followed by cyclization mediated by potassium carbonate was utilized to prepare the N-

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<sup>(8)</sup> Fearon, K.; Spaltenstein, A.; Hopkins, P. B.; Gelb, M. H. J. Med. Chem. 1987, 30, 1617. Thaisrivongs, S.; Pals, D. T.; Kati, W. M.; Thomasco, L. M. J. Med. Chem. 1985, 28, 1553. Gelb, M. H.; Svaren, J. P.; Abeles, R. H. Biochemistry 1985, 24, 1813. Rich, D. H.; Bernatowicz, M. S.; Schmidt, P. G. J. Am. Chem. Soc. 1982, 104, 3536. Bartlett, P. A.; Kezer, W. B. J. Am. Chem. Soc. 1984, 106, 4282. Bock, M. G.; DiPardo, R. M.; Evans, B. E.; Rittle, K. E.; Boger, J. S.; Freidinger, R. M.; Veber, D. F. J. Chem. Soc., Chem. Commun. 1985, 109.

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<sup>(12)</sup> Andrew, R. G.; Conrow, R. E.; Elliott, J. D.; Johnson, W. S.; Ramezani, S. Tetrahedron Lett. 1987, 28, 6535. Harris, B. D.; Bhat, K. L.; Joullie, M. Tetrahedron Lett. 1987, 28, 2837. Woo, P. W. K. Tetrahedron Lett. 1985, 26, 2973 and pertinent references cited therein. (13) Rich, D. H.; Sun, E. T.; Bopari, A. S. J. Org. Chem. 1978, 43, 3624.

 <sup>(14)</sup> Mitsunobu, O. Synthesis 1981, 1.
 (15) Mattingly, P. G.; Kerwin, J. F.; Miller, M. J. J. Am. Chem. Soc.
 1979, 101, 3983. Miller, M. J.; Mattingly, P. G.; Morrison, M. A.; Kerwin, J. F. J. Am. Chem. Soc. 1980, 102, 7026.

<sup>(16)</sup> Alternatively, carbon tetrachloride/triphenylphosphine<sup>14</sup> or the two-step sequence mesylation/cyclization<sup>17</sup> was employed.
(17) Floyd, D. M.; Fritz, A. W.; Plusec, J.; Weaver, E. R.; Cimarusti, C. M. J. Org. Chem. 1982, 47, 5160. Evans, D. A.; Sjogren, E. B. Tetra-ballor, J. 1982, 1982, 1982, 1983. hedron Lett. 1986, 27, 4961.

methoxylactam 7b. This intermediate was subjected to the same reduction/hydrolysis/protection protocol to provide the desired 10b as a white crystalline product. We had, to this point, relied on literature precedent in the assignment of the absolute stereochemistry of 9a and 9b. Carbon-13 NMR spectra on both 6a and 7a had shown that a single diastereomer (7a) had been prepared from a diastereomerically pure starting material (6a). Miller's<sup>15</sup> studies had reported that these intramolecular reactions proceeded with inversion of configuration at the reaction center. The acids 9a and 9b were converted to the lactams 12a and 12b, respectively, as shown in Scheme II, to verify that inversion had occurred and the structural assignments presented in Scheme I were correct.

The free acids were converted into their methyl esters<sup>18</sup> and cyclized to the lactams by utilizing tert-butyldimethylsilyl trifluoromethanesulfonate followed by an aqueous bicarbonate workup. The cyclization most likely proceeded via an N-silyl intermediate that closed intramolecularly under the catalysis of the excess triflate. High field (500 MHz) NOE studies were then carried out to determine the relationship between the protons at carbons 4 and 5 in 12a and 12b. Irradiation of proton H4 in 12a resulted in NOE's of 7.1% to H5 and 8.6% to H3a, while irradiation of H5 resulted in NOE's of 11.4% to H4 and 1.9% to H3a. Irradiation of H4 in 12b gave only a 1.3% NOE to H5 and irradiation of H5 gave a 1.8% NOE to H4. This evidence verified the cis relationship of protons H5 and H4 in 12a (trans in 12b), the integrity of the inversion during intramolecular cyclization, and the absolute stereochemistry of the structures assigned to the products 9a (3S,4S) and **9b** (3R,4S).

The initial goal of this effort was the development of a stereospecific synthesis of the statine isosteres 3 and 4, which were suitably protected for further synthetic elaboration. We have accomplished this objective with the synthesis of the 3-aminodeoxystatine derivatives 9a and 9b. Further research directed toward the utilization of these intermediates in inhibition studies of the biologically important aspartyl proteinases has been published elsewhere. 1.19

## **Experimental Section**

Mass spectra, infrared spectra, optical rotations, combustion analyses, and high field  $^1H$  NMR spectra at 200 MHz and 500 MHz were obtained by the Physical and Analytical Chemistry Department of The Upjohn Company. Other  $^1H$  NMR spectra were determined on a Varian CFT-20 spectrophotometer at 80 MHz and chemical shifts are reported as  $\delta$  units relative to tetrmethylsilane. Melting points are uncorrected.

Thin-layer chromatography was conducted with Analtech 0.25-mm glass plates precoated with silica gel GF. For column chromatography, E. Merck silica gel 60, 230-400 mesh, or E. Merck prepacked Lobar columns were used. All solvents for chromatography were Burdick and Jackson reagent grade distilled in glass.

Tetrahydrofuran was distilled under argon from sodium metal in the presence of benzophenone. Dichloromethane was distilled from calcium hydride. Diethyl phosphorocyanidate<sup>20</sup> was freshly distilled before use.

(3R,4S)-3-Hydroxy-4-[[(1,1-dimethylethoxy)carbonyl]-amino]-6-methylheptanoic acid (5a) and its 3S,4S isomer 5b were prepared by the method of Rich et al.<sup>13</sup>

(1S,2R)-[2-Hydroxy-4-(methoxyamino)-1-(2-methylpropyl)-4-oxobutyl]carbamic Acid 1,1-Dimethylethyl Ester (6a). A solution of 5a (5.0 g, 18.2 mmol), O-methylhydroxylamine

hydrochloride (1.67 g, 20.0 mmol), and diethyl phosphorocyanidate (3.26 g, 20.0 mmol) in dichloromethane (100 mL) was stirred at 0 °C under argon and triethylamine (5.31 mL, 38.2 mmol) added via syringe over a period of 5 min. The reaction mixture was stirred at 0 °C for 1 h, poured into water, washed with brine, dried (anhydrous sodium sulfate), and concentrated to give a crude, crystalline product. Recrystallization of the residue from diethyl ether gave 3.2 g (58%) of 6a as a fluffy white solid, mp 134-5 °C (from ether):  $[\alpha]^{25}_D$  -13.5° (c 0.892, MeOH); IR (cm<sup>-1</sup>, mull) 3334, 3270, 3040, 1680, 1668, 1535, 1389, 1174; <sup>1</sup>H NMR (80 MHz, CDCl<sub>3</sub>)  $\delta$  9.42 (br s, 1 H), 4.57 (d, J = 8 Hz, 1 H), 3.78–3.48 (m, 1 H), 3.77 (s, 3 H), 3.48-2.93 (m, 2 H), 2.30 (dd, J = 1.4, 3.5 Hz, 2 H), 1.60-0.91 (m, 3 H), 1.44 (s, 9 H), 0.92 (d, J = 6.0 Hz, 3 H), 0.90 (d, J = 6.0 Hz, 3 H); MS (CI, isobutane), m/z (relative intensity)  $305 [M + H]^+ (100)$ , 249 (60), 609 (32), 306 (18), 189(13). Anal. Calcd for C<sub>14</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>: C, 55.14; H, 9.20; N, 9.20. Found: C, 54.94; H, 9.33; N, 9.14.

(1S,2S)-[2-Hydroxy-4-(methoxyamino)-1-(2-methylpropyl)-4-oxobutyl]carbamic Acid 1,1-Dimethylethyl Ester (6b). The synthesis of 6b was carried out as described above for 6a starting with 5b (2.50 g, 9.1 mmol). Recrystallization from ethyl acetate/hexane provided 6b (1.85 g, 67%), mp 92.5–94.5 °C: IR (mull, cm<sup>-1</sup>) 3280, 3203, 1685, 1664, 1443, 1399, 1341, 1171; <sup>1</sup>H NMR (80 MHz, CDCl<sub>3</sub>) δ 8.20 (br s, 1 H), 4.78 (d, J = 10 Hz, 1 H), 4.16–3.79 (m, 1 H), 3.75 (s, 3 H), 3.75–3.35 (m, 1 H), 2.27 (d, J = 6.3 Hz, 2 H), 1.88–1.09 (m, 3 H), 1.44 (s, 9 H), 0.90 (d, J = 5.8 Hz, 6 H); MS (EI), m/z (relative intensity) 86 (100), 130 (87), 57 (85), 118 (51), 140 (30). Anal. Calcd for C<sub>14</sub>H<sub>28</sub>N<sub>2</sub>O<sub>5</sub>: C, 55.14; H, 9.20; N, 9.20. Found: C, 55.37; H, 9.37; N, 9.24.

(1S,2S)-[1-(1-Methoxy-4-oxo-2-azetidinyl)-3-methylbutyl]carbamic Acid 1,1-Dimethylethyl Ester (7a). A solution of 6a (304 mg, 1.0 mmol) and triphenylphosphine (524 mg, 2.0 mmol) in dry THF (5 mL) was stirred at room temperature under argon and diethyl azodicarboxylate (0.32 mL, 4.0 mmol) added via syringe. The reaction mixture was stirred for 30 min, concentrated in vacuo, and chromatographed on silica gel (elution with ethyl acetate/hexane 1:2) to deliver 200 mg (70%) of 7a as a white crystalline solid, mp 96.5-97.5 °C (hexane/diethyl ether):  $[\alpha]^{25}_{D}$  -76.2° (c 0.795, MeOH); IR (mull, cm<sup>-1</sup>) 3348, 2951, 1797, 1792, 1682, 1537, 1250, 1176; <sup>1</sup>H NMR (80 MHz, CDCl<sub>3</sub>) δ 4.55-4.17 (m, 1 H), 4.17-3.60 (m, 2 H), 3.78 (s, 3 H), 2.60 (dd, J = 12.6, 4.7 Hz, 1 H), 2.56 (dd, J = 12.6, 2.8 Hz, 1 H), 1.88–1.09 (m, 3 H), 1.43 (s, 9 H), 0.94 (d, J = 6.2 Hz, 6 H); MS (CI, NH<sub>3</sub>),m/z (relative intensity) 248 (100), 304 [M + NH<sub>4</sub>]<sup>+</sup> (95), 204 (29), 266 (21), 305 (16). Anal. Calcd for  $C_{14}H_{26}N_2O_4$ : C, 58.72; H, 9.15; N, 9.78. Found: C, 58.85; H, 9.29; H, 9.80.

(1S,2R)-[1-(1-Methoxy-4-oxo-2-azetidinyl)-3-methylbutyl]carbamic Acid 1,1-Dimethylethyl Ester (7b). A cold (0 °C) solution of 6b (1.20 g, 3.95 mmol) in dry pyridine (10 mL) was stirred under argon and methanesulfonyl chloride (0.62 mL, 8.0 mmol) added via syringe. The reaction mixture was stirred at 0 °C for 2 h, poured into brine, and extracted with ethyl acetate. The extracts were combined, dried with anhydrous sodium sulfate, and concentrated. The red residue so obtained was dissolved in acetone (20 mL), anhydrous potassium carbonate (1.0 g, 7.2 mmol) was added, and the slurry was refluxed for 72 h. The solution was gooled, diluted with ethyl acetate, filtered through a medium porosity scintered glass funnel, and concentrated. Chromatography of the residue on silica gel (Merck Lobar B, elution with ethyl acetate/hexane 1:3) gave 680 mg (60%) of 7b as a slightly yellow oil: IR (film, cm<sup>-1</sup>) 3342, 2958, 1777, 1708, 1691, 1526, 1366, 1250, 1170; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  4.46 (d, J = 10 Hz, 1 H), 4.14-3.95 (m, 1 H), 3.90-3.82 (m, 1 H), 3.83 (s, 3 H), 2.77 (dd, J = 14.3, 5.4 Hz, 1 H), 2.52 (dd, J = 14.3, 3.0 Hz, 1 H), 1.73 (sep,J = 7.2 Hz, 1 H, 1.45 (s, 9 H), 1.35-1.20 (m, 2 H), 0.96 (d, J = 1.35 (m, 2 H), 0.96 (d, J6.6 Hz, 3 H), 0.95 (d, J = 6.8 Hz, 3 H); MS (FAB), m/z (relative intensity) 187 (100), 57 (44), 231 (38), 154 (18), 140 (13), 128 (12); exact mass calcd for C<sub>14</sub>H<sub>26</sub>N<sub>2</sub>O<sub>4</sub> 287.1971, obsd 287.1964.

(1S,2S)-[1-(4-Oxo-2-azetidinyl)-3-methylbutyl]carbamic Acid 1,1-Dimethylethyl Ester (8a). Sodium (75 mg, 3.1 mmol) was added to a cold (-78 °C) solution of dry ammonia (20 mL), and the mixture was stirred for 15 min until all the metal had dissolved. A solution of 7a (200 mg, 0.7 mmol) in dry THF (5 mL) was slowly added, and the reaction mixture was allowed to warm to reflux (-33 °C). After 30 min, solid ammonium chloride (200 mg, 3.7 mmol) was added, and the resulting solution was

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<sup>(20)</sup> Yamada, S.; Kasai, Y. Tetrahedron Lett. 1973, 1595.

allowed to warm to room temperature over a period of 2 h. The residue was diluted with ethyl acetate (50 mL), washed with water, dried (anhydrous sodium sulfate), and concentrated to deliver 180 mg (100%) of 8a as a clear colorless oil:  $[\alpha]^{25}_{\rm D}$ –26.6° (c 1.09, MeOH); IR (film, cm<sup>-1</sup>) 3317, 2956, 1761, 1707, 1685, 1535, 1367, 1252; <sup>1</sup>H NMR (80 MHz, CdCl<sub>3</sub>)  $\delta$  6.15 (br s, 1 H), 4.53 (d, J = 9.3 Hz, 1 H), 4.03–3.53 (m, 2 H), 2.89 (dd, J = 2.3, 4.6 Hz, 1 H), 2.77 (d, J = 1.7 Hz, 1 H), 1.90–1.58 (m, 1 H), 1.44–1.11 (m, 2 H), 1.44 (s, 9 H), 0.93 (d, J = 6.3 Hz, 6 H); MS (CI, NH<sub>3</sub>), m/z (relative intensity) 274 [M + NH<sub>4</sub>]<sup>+</sup> (100), 69 (43), 124 (16), 218 (13), 94 (6); exact mass calcd for  $C_{13}H_{24}N_2O_3$  256.1787, obsd 256.1794.

(1S,2R)-[1-(4-Oxo-2-azetidinyl)-3-methylbutyl]carbamic Acid 1,1-Dimethylethyl Ester (8b). The synthesis of 8b was carried out as described above for 8a, starting with 7b (4.70 g, 16.4 mmol). Chromatography on silica gel (elution with ethyl acetate/hexane 2:1) provided 8b (3.76 g, 90%) as a white crystalline solid, mp 135.5–137 °C (from ethyl acetate/hexane):  $[\alpha]^{24}_{D}$ -38.3° (c 1.00, MeOH); IR (mull, cm<sup>-1</sup>) 3381, 3198, 1748, 1718, 1680, 1512, 1252, 1162; <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  5.96 (br s, 1 H), 4.40 (d, J = 9.0 Hz, 1 H), 3.94 (m, 1 H), 3.63 (dt, J = 2.6, 4.6 Hz, 1 H), 2.92 (dd, J = 4.7, 15.4 Hz, 1 H), 2.70 (m, 1 H), 1.84–1.62 (m, 1 H), 1.46 (s, 9 H), 1.30–1.18 (m, 2 H), 0.97 (d, J = 6.7 Hz, 3 H), 0.95 (d, J = 6.2 Hz, 3 H); MS (EI), m/z (relative intensity) 57 (100), 86 (89), 130 (57), 186 (32), 71 (14). Anal. Calcd for  $C_{13}H_{24}N_{2}O_{3}$ : C, 60.91; N, 9.44; N, 10.93. Found: C, 60.71; H, 9.30, N, 10.81.

(3S,4S)-4-[[(1,1-Dimethylethoxy)carbonyl]amino]-6methyl-3-[[(phenylmethoxy)carbonyl]amino]heptanoic Acid (9a). A solution of 8a (466 mg, 1.8 mmol) and potassium hydroxide (10 mL of 1 N solution, 10 mmol) in methanol (10 mL) was stirred at room temperature for 3 h, cooled to 0 °C, neutralized with 6 N HCl, and concentrated. The residue was dissolved in DMF (5 mL), potassium carbonate added (10 mL of 1 M solution, 10 mmol), and the solution cooled to 0 °C. CBZCl (0.64 mL, 4.5 mmol) was added, the ice bath removed after 4 h, and the solution stirred at room temperature for 48 h. The reaction mixture was washed with diethyl ether, acidified to pH 2 with 6 N HCl, and extracted with dichloromethane. The extracts were combined, dried with anhydrous magnesium sulfate, and concentrated to give 750 mg (100%) of 9a as a slightly yellow oil (crystalline analytical sample obtained upon acidification of the DCHA salt, mp 133-4.5 °C (lit. 10 mp 128-130 °C):  $[\alpha]^{25}_{D}$  -43.5 ° (c 0.575, MeOH) (lit. 10  $[\alpha]^{25}_{D}$  -46.1 ° (c 0.5, MeOH)); IR (film cm<sup>-1</sup>) 3310, 2940, 1690, 1500, 1230, 1150; <sup>1</sup>H NMR (80 MHz, CDCl<sub>3</sub>) δ 9.24 (br s, 1 H), 7.30 (s, 5 H), 6.86-5.36 (m, 1 H), 5.07 (s, 2 H), 5.08-4.50 (m, 1 H), 4.22-3.47 (m, 2 H), 2.64-2.37 (m, 2 H), 1.70-0.79 (m, 3 H), 1.39 (s, 9 H), 0.87 (d, J = 6.0 Hz, 6 H); MS (EI), m/z (relative intensity) 91 (100), 86 (57), 54 (43), 186 (21), 136 (14), 408 (M<sup>+</sup>, 4). Anal. Calcd for  $C_{21}H_{32}N_2O_6$ : C, 61.75; H, 7.90; N, 6.86. Found: C, 61.76; H, 8.06, N, 6.87.

(3R,4S)-4-[[(1,1-Dimethylethoxy)carbonyl]amino]-6-methyl-3-[[(phenylmethoxy)carbonyl]amino]heptanoic Acid (9b). The synthesis of 9b was carried out as described above for 9a starting with 8b (2.16 g, 8.4 mmol) to provide 9b (3.40 g, 100%) as an off-white powder (crystalline analytical sample obtained upon acidification of the DCHA salt, mp 198–200 °C (lit. 10 mp 203–5 °C:  $[\alpha]^{25}_{\rm D}-3.3^{\circ}$  (c 1.0, MeOH) (lit.  $[\alpha]^{25}_{\rm D}-3.6^{\circ}$  (c 0.5, MeOH)); 1H NMR (80 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD 1:1)  $\delta$  7.32 (s, 5 H), 5.94 (m, 1 H), 5.09 (s, 2 H), 5.19–4.82 (m, 1 H), 3.79 (m, 2 H), 2.51 (m, 2 H), 1.42 (s, 9 H), 1.61–1.10 (m, 3 H), 0.92 (d, J = 6.1 Hz, 3 H), 0.88 (d, J = 6.2 Hz, 3 H); MS (EI), m/z (relative intensity) 57 (100), 86 (66), 91 (66), 130 (42), 139 (17), 186 (17).

(3S,4S)-4-[[(1,1-Dimethylethoxy)carbonyl]amino]-6-methyl-3-[[(phenylmethoxy)carbonyl]amino]heptanoic Acid with N-Cyclohexylcyclohexanamine (10a). A solution of 9a (1.44 g, 3.52 mmol) was stirred at room temperature in diethyl ether (20 mL) under argon and freshly distilled dicyclohexylamine (905 mg, 5.0 mmol) added via syringe. The resulting solution was stirred at room temperature overnight and the product filtered and dried to deliver 927 mg (45%) of 10a as a white crystalline solid, mp 150–152 °C (ethyl acetate):  $[\alpha]^{26}_{\rm D}$  –28.5° (c 0.789, MeOH); IR (mull, cm<sup>-1</sup>) 3324, 1923, 1723, 1713, 1538, 1451, 1255; MS (CI, NH<sub>3</sub>), m/z (relative intensity) 426 [M + NH<sub>4</sub>]<sup>+</sup> (100), 275 (54), 182 (25), 427 (24), 308 (13). Anal. Calcd for  $C_{33}H_{55}N_3O_6$ : C, 67.20; H, 9.40; N, 7.13. Found: C, 66.93; H, 9.59; N, 6.94.

(3S,4R)-4-[[(1,1-Dimethylethoxy)carbonyl]amino]-6-methyl-3-[[(phenylmethoxy)carbonyl]amino]heptanoic Acid with N-Cyclohexylcyclohexanamine (10b). The synthesis of 10b was carried out in ethyl acetate as described above for 10a starting with 9b (95 mg, 0.23 mmol) to give 10b (96 mg, 71%) as an analytically pure solid, mp 166–167.5 °C (ethyl acetate): IR (mull, cm<sup>-1</sup>) 3427, 3316, 1710, 1681, 1639, 1550, 1515, 1238; MS (CI, NH<sub>3</sub>), m/z (relative intensity) 275 (100), 426 [M + NH<sub>4</sub>]<sup>+</sup> (68), 382 (25), 308 (19), 427 (18). Anal. Calcd for  $C_{33}H_{55}N_3O_6$ : C, 67.20; H, 9.40; N, 7.13. Found: C, 67.19; H, 9.44; N, 7.04.

Methyl (3S,4S)-4-[[(1,1-Dimethylethoxy)carbonyl]amino]-6-methyl-3-[[(phenylmethoxy)carbonyl]amino]heptanoate (11a). A solution of 9a (17 mg, 0.042 mmol), methyl iodide (8.2 mg, 0.058 mmol), and DBU (9.5 mg, 0.067 mmol) in acetonitrile (1 mL) was stirred at room temperature under argon for 72 h. The reaction mixture was concentrated, passed through a Waters Sep-Pak (silica, elution with 9:1 hexane/ethyl acetate), and evaporated to give 12 mg (68%) of 11a as a clear colorless oil:  $[\alpha]^{25}_{\rm D}$  -39.5° (c 0.597, MeOH); IR (mull, cm<sup>-1</sup>) 3357, 1741, 1697, 1518, 1249, 1173; <sup>1</sup>H NMR (80 MHz, CDCl<sub>3</sub>) & 7.32 (s, 5 H), 5.42 (m, 1 H), 5.08 (s, 2 H), 4.48 (m, 1 H), 4.23-3.53 (m, 2 H), 3.64 (s, 3 H), 2.56 (m, 2 H), 1.83-1.07 (m, 3 H), 1.40 (s, 9 H), 0.90 (d, J = 6.2 Hz, 3 H), 0.88 (d, J = 6.2 Hz, 3 H); MS (FAB), m/z (relative intensity) 423 [M + H]<sup>+</sup>; exact mass calcd for  $C_{22}H_{34}N_2O_6$  423.2495, obsd 423.2508.

Methyl (3R,4S)-4-[[(1,1-Dimethylethoxy)carbonyl]-amino]-6-methyl-3-[[(phenylmethoxy)carbonyl]amino]heptanoate (11b). The synthesis of 11b was carried out as described above for 11a starting with 9b (20 mg, 0.049 mmol) to provide 11b (19 mg, 82%) as a white crystalline solid:  $^1$ H NMR (80 MHz, CDCl<sub>3</sub>) δ 7.33 (s, 5 H), 5.61 (m, 1 H), 5.10 (s, 2 H), 4.38 (m, 1 H), 4.16–3.50 (m, 2 H), 3.64 (s, 3 H), 2.54 (d, J=5.2 Hz, 2 H), 1.80–1.23 (m, 3 H), 1.42 (s, 9 H), 0.90 (d, J=6.1 Hz, 3 H), 0.88 (d, J=6.0 Hz, 3 H); MS (FAB), m/z (relative intensity) 423 [M + H]<sup>+</sup>; exact mass calcd for  $C_{22}H_{34}N_2O_6$  423.2495, obsd 423.2528.

(4S,5S)-4-[[(Phenylmethoxy)carbonyl]amino]-5-(2'methylpropyl)-2-oxopyrrolidine (12a). A solution of 11a (12 mg, 0.028 mmol) and tert-butyldimethylsilyl trifluoromethanesulfonate (0.2 mL, 0.87 mmol) in dichloromethane (1 mL) was stirred at room temperature under argon for 4 h. The reaction mixture was quenched with saturated sodium carbonate solution, extracted with dichloromethane, dried with anhydrous sodium sulfate, and concentrated. Chromatography of the residue on silica gel (Merck Lobar A, elution with 2.5% MeOH/dichloromethane) gave 5.2 mg (63%) of 12a as a clear colorless oil which crystallized on standing: <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD) δ 7.38 (m, 5 H), 5.06 (AB q, J = 11.9 Hz, 2 H), 4.44 (m, 1 H), 3.80 (m, 1 H), 2.98 (m, 1 H), 2.61 (dd, J = 17.3, 7.9 Hz, 1 H), 2.18 (dd, J = 17.3,4.8 Hz, 1 H), 1.57 (m, 1 H), 1.31-1.20 (m, 2 H), 0.89 (d, J = 6.5Hz, 3 H), 0.85 (d, J = 6.5 Hz, 3 H), other exchangeable protons not observed; MS (FAB), m/z (relative intensity) 291 [M + H]<sup>+</sup>, exact mass calcd for C<sub>16</sub>H<sub>22</sub>N<sub>2</sub>O<sub>3</sub> 291.1709, obsd 291.1718.

(4R,5S)-4-[[(Phenylmethoxy)carbonyl]amino]-5-(2'-methylpropyl)-2-oxopyrrolidine (12b). The synthesis of 12b was carried out as described above for 12a starting with 11b (11 mg, 0.026 mmol) to provide 12b (6.5 mg, 86%) as a clear colorless oil which crystallized on standing:  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>/CD<sub>3</sub>OD) δ 7.34 (m, 5 H), 6.35 (s, 1 H), 5.40 (s, 1 H), 5.10 (AB q, J = 13.0 Hz, 2 H), 4.05 (m, 1 H), 3.95 (m, 1 H), 3.48 (m, 1 H), 2.75 (dd, J = 17.6, 8.4 Hz, 1 H), 2.19 (dd, J = 17.6, 5.2 Hz, 1 H), 1.67 (m, 1 H), 1.58 (m, 1 H), 1.49 (m, 1 H), 0.94 (d, J = 6.5 Hz, 3 H), 0.92 (d, J = 6.5 Hz, 3 H); MS (FAB), m/z (relative intensity) 291 [M + H]<sup>+</sup>, exact mass calcd for  $C_{16}H_{22}N_2O_3$  291.1709, obsd 291.1724.

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